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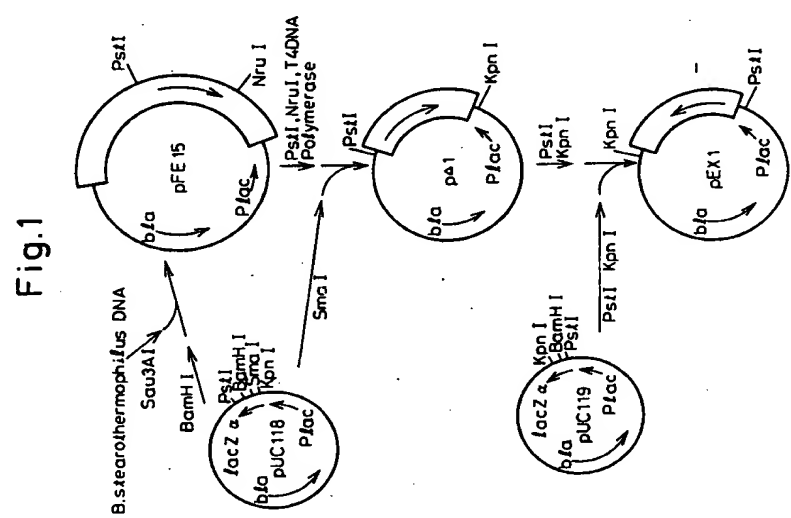
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**Farnesyl pyrophosphate synthetase and DNA sequence encoding the same.**

A DNA sequence that encodes a stable farnesyl pyrophosphate synthetase and the use thereof are provided. DNA sequence that encodes a farnesyl pyrophosphate synthetase originating from *Bacillus stearothermophilus* is provided. By selecting *Bacillus stearothermophilus* as the gene origin of the synthetase, a production system for the synthetase, a production system for the synthetase particularly having thermal stability, can be constructed.



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## BACKGROUND OF THE INVENTION

## 1. Field of the Invention

5 The present invention relates to a bacterial farnesyl pyrophosphate synthetase. More specifically, the invention relates to a DNA sequence encoding a farnesyl pyrophosphate synthetase active substance originating from Bacillus stearothermophilus, a transformant carrying such a DNA sequence and a process for producing a farnesyl pyrophosphate synthetase active substance and farnesyl diphosphate. In accordance with the invention, accordingly, it is capable of producing the farnesyl pyrophosphate synthetase with  
10 high efficiency by the transformant.

## 2. Description of Related Art

15 Farnesyl pyrophosphate (hereinafter sometimes abbreviated as "FPP") is an important intermediate for bio-synthesizing carotenoids, cholesterol, isoprenoids such as gum, and the like, and has been known to be formed by the condensation of isopentenyl pyrophosphate with 3,3-dimethylallyl diphosphate and it has also been known that its synthetase exist in many living organisms. Accordingly, it is assumed to be useful when the gene of the FPP synthetase is introduced in an appropriate host, and an isoprenoid is expected to be produced on a larger scale.

20 Base on the above attempts have been to study genes encoding a microorganism originated FPP synthetase and manipulations thereof and to produce the synthetase, but only enzymes originating from Escherichia coli have been known (J. Biochem. 108, page 995-100 (1990)). Moreover, the FPP synthetase originating from Escherichia coli is considerably unstable, and its activity is rapidly inactivated, for example, at 50°C. Furthermore, the synthetase coding genes originating from Escherichia coli are only introduced in  
25 another Escherichia coli, and no example of an attempt to introduce the same into any other microorganism has yet been described in any known literature.

## SUMMARY OF THE INVENTION

30 In utilizing the FPP synthetase for practically producing FPP, it is insufficient to use the synthetase originating from Escherichia coli as described above and, therefore, the need for providing an FPP synthetase, in particular, one having thermal stability, would still exist. Accordingly, the object of the present invention is to provide a DNA sequence encoding a thermalstable FPP synthetase and to construct a thermalstable FPP production system using such a DNA sequence.

35 When the present inventors researched farnesyl pyrophosphate synthetase of a wide variety of microorganisms in order to solve the above problem, it has been found that Bacillus stearothermophilus, which has been generally known to grow at a high temperature and to produce various thermally stable enzymes as a rule, also produces an FPP synthetase and has further succeeded in expressing a corresponding gene by a genetically engineered technique, thereby achieving this invention.

40 Consequently, the above problem can be solved by a DNA sequence that encodes a farnesyl pyrophosphate synthetase originating from Bacillus stearothermophilus, a recombinant vector carrying the DNA sequence, and a recombinant microorganism cell having a gene transferred therein by the recombinant vector.

45 According to the present invention, there is also provided a process for producing a farnesyl pyrophosphate synthetase active substance that comprises culturing the microorganisms in a nutrient medium to produce and accumulate the synthetase active substance in the cultures, and isolating the synthetase therefrom.

50 According to the present invention, there is also provided use of the farnesyl pyrophosphate synthetase, i.e., a process for producing a farnesyl pyrophosphate that comprises condensing a 3,3-dimethylallyl pyrophosphate or geranyl pyrophosphate with isopentenyl pyrophosphate in the presence of a farnesyl pyrophosphate synthetase active substance to form the farnesyl pyrophosphate.

Thus, wide use of a farnesyl pyrophosphate synthetase active substance useful for producing a farnesyl pyrophosphate is possible.

## 55 BRIEF DESCRIPTION OF DRAWINGS

The present invention will be illustrated by referring to the drawings attached to the specification. Figure 1 is a flow chart of the construction of a recombinant plasmid according to the invention, and

Figure 2 is a restriction enzymes map of the plasmid pFE 15 according to the invention.

#### DETAIL DESCRIPTION

5 The term "DNA sequence which encodes a farnesyl pyrophosphate synthetase" is herein intended to mean all enzymes coding DNA units, that can express the gene encoding the synthetase, when they are incorporated into an appropriate expression vector, and is used in the conception of encompassing all enzymes encoding substantially equal enzyme active substances. Examples of them, specifically, include all DNA sequences encoding the amino acid sequences depicted on Sequence No. 1 of the Sequence  
10 Listing. Also included herein are enzymes that encode such DNA sequences and, at the same time, encode additional amino acids as long as the proteins produced by the expression have the above mentioned activity (for example, fused-proteins). Specific examples of such DNA sequences are DNA sequences as depicted on Sequence No. 1 (SEQ ID NO:1) of the Sequence Listing.

The DNA sequence of the present invention may be prepared from a chromosome of *Bacillus*  
15 *stearothermophilus*, which is available from various Institutes for the Deposit of Microorganisms, by any method known per se, the details of which are described later.

The invention also provides recombinant vector comprising the above-mentioned DNA sequence. Such a recombinant vector also contains a DNA sequence, and has a function of regulating the expression of gene encoding the FPP synthetase.

20 For example, in the case of using *Escherichia coli* in the host, it has been known that there are functions for regulating the expression of genes, such as the stage of transcription from DNA to mRNA and the stage for translation from mRNA to a protein. In addition to naturally occurring sequences, e.g., lac, trp, bla, lpp, P<sub>L</sub>, P<sub>R</sub>, tet, T3, T7, etc, mutants thereof, e.g., lac UV5, and sequences in which naturally occurring promoter coding sequences are artificially fused, e.g., tac, trc, etc., have been known as the promoter  
25 sequences for regulating the synthesis of mRNA, and they can be used in the invention. In the sequences which regulate the capability of translation of a protein from mRNA, the distance between the ribosome binding site sequence (GAGG and similar sequences) and the initiation codon "ATG" has already been known to be important. It is also well known that a vector containing a terminator that directs termination of the transcription to 3'-end, e.g., a vector containing rrnBT<sub>1</sub>T<sub>2</sub>, which is available from Pharmacia, affects the  
30 efficiency of synthesizing a protein in a recombinant.

In the DNA disclosed by this invention, the gene of FPP synthetase starts from GTG encoding rare initiation codon, but it can easily be deduced that the synthesis efficiency would be increased if it starts from ATG, which is a usual initiation codon for synthesizing a protein.

For vectors that can be used for preparing the recombinant vector of the invention, a commercially  
35 available vector itself may be useful and various vectors that are also derived according to the object may be useful. For example, such vectors include pBR322 pBR327, pKK223-3, pKK233-2, pTrc991, and the like, which possess a replicon originating from pMB1; pUC18, pUC19, pUC118, pUC119, pHSG298, pHSG396, and the like, which have been modified so as to increase the copy number; pACYC117, pACYC118, and the like, which possess a replicon originating from p15A; and plasmids derived from pSC101, ColE1 or R1, and  
40 one derived from F factor, and the like. Alternatively, other than the plasmid, it is also possible to incorporate a gene into a host cell by means of a virus vector or a transposon such as  $\lambda$  phage or M13 phage.

For the gene incorporated into microorganisms other than *Escherichia coli*, there is a gene incorporated into the microorganisms belonging to the genus *Bacillus* by means of pUB110 (sold by Sigma) or  
45 pPHY300PLK (sold by Takara Shuzo Co., Ltd.). For these vectors, there are descriptions in Molecular cloning (J. Sambrook, E. F. Fritsch, and T. Maniatis Ed., pressed by Cold Spring Harbor Laboratory Press) and Cloning Vector (P. H. Pouwels, B. E. Enger-Valk and W. J. Brammar Ed., pressed by Elsevier), and catalogues of the producers. Particularly, pTrc 99A (sold by Pharmacia), which not only has a resistant gene to ampicillin, which is a selected marker, but also has P<sub>trc</sub> and lacI<sup>q</sup> as a promoter and a regulation gene,  
50 respectively, the sequence of "AGGA", as the ribosome-binding site, and rrnBT<sub>1</sub>T<sub>2</sub>, as the terminator, and functions to regulate the expression of an FPP synthetase, can be mentioned as a preferable vector.

The insertion of the DNA fragment that encodes an FPP synthetase and, if necessary, a DNA fragment possessing the function of regulating the expression of an FPP synthetase into these vectors can be carried out by a known method using appropriate restriction enzymes and ligase. More specifically, this is  
55 preferably conducted according to the method described below. An example of the plasmid produced as described above is pEX1. This plasmid can be obtained from *Escherichia coli* JM109 (pEX1), which has been internationally, under the Budapest Treaty, deposited at the Fermentation Reserach Institute, Agency of Industrial Science and Technology, as FERM BP-3581, on September 26, 1991, by the method known

per se.

As the microorganisms that can be gene-incorporated, microorganisms belonging to Escherichia coli, Bacillus, and the like, can be utilized. The transformation or transduction thereof may also be conducted by a conventional method such as a  $\text{CaCl}_2$  method or protoplast method described, for example, in Molecular cloning (J. Sambrook, E. F. Fritsch, and T. Maniatis Ed, pressed by Cold spring Harbor Laboratory Press), DNA Cloning Vol. I-III (D. M. Glover ed., pressed by IRL PRESS), and the like. As a representative transformant thus obtained, the above-mentioned JM109 (pEX1) (also described as pEX 1/JM109) can be considered.

When being cultured in a nutrient medium usually used in the culture of Escherichia coli, these transformants or recombinant accumulate farnesyl pyrophosphate in their bacterial cells, in the case of using Escherichia coli as the host. For example, as the nutrient medium, any of the synthesized media and natural media containing carbon sources, nitrogen sources, and mineral substances known per se may be used. As carbon sources, carbohydrates, such as glycerol glucose, glycerine, fructose, sucrose, maltose, starches, starch hydrolyzed liquid syrups, or the like can be used. The amount used is preferably about 0.1 to 5.0%.

As nitrogen sources, various inorganic and organic ammonium salts, such as ammonia, ammonium chloride, ammonium phosphate, ammonium sulfate, and naturally originating nitrogen sources, such as amino acids, meat extracts, yeast extracts, corn steep liquors, casein hydrolyzed products, defatted soybean powder and digested products thereof, and the like may be used. The naturally originating nitrogen sources, in many cases, also serve as carbon sources, in addition to the nitrogen sources.

As the mineral substances, potassium dihydrogenphosphate, dipotassium hydrogenphosphate, magnesium sulfate, sodium chloride, copper sulfate, ferrous sulfate, manganese chloride, cobalt chloride, ammonium molybdate, boric acid, and the like can be used.

The culture is preferably carried out under aerobic conditions, such as by shaking culture or aeration culture. The culture temperature is suitably 25 - 37°C, and the pH level during the culture is preferably maintained around the neutral pH level. Usually, the period of the culture is approximately 24 - 72 hours.

The collection of FPP from the bacterial cells can be performed by treating the bacterial cells either physically or in an environment with the presence of an appropriate lytic enzyme to be subjected to lysis, and then removing cell debris followed by general methods for the isolation and purification of an enzyme.

As the lytic enzyme, the use of lysozyme is preferable, and as the physical treatment, the use of sonication is preferable. By a thermal treatment at approximately 55°C, many proteins originating from Escherichia coli can be removed out as insoluble precipitates. The isolation and purification of the enzyme may be carried out using various kinds of chromatography such as gel permeation, ion-exchanging, hydrophobic, reverse phase, and affinity chromatography or using superfiltration singly or in combination. As a stabilizer for stabilizing the desired enzyme throughout the isolation and purification stages, for example, reducing agents such as  $\beta$ -mercaptoethanol and dithiothreitol, agents for protecting protease such as PMS and BSA, metal ions such as magnesium may coexist in the liquid to be treated.

Since the activity of the above-mentioned FPP synthetase can be measured, for example, as follows, the isolation and purification are recommended to be carried out while monitoring the enzyme activity using a reaction liquid for assay described later in Example 1 c).

Furthermore, the assay system, as such or in a modified condition, can be utilized for producing the farnesyl pyrophosphate via condensation of geranyl pyrophosphate with isopentenyl pyrophosphate in the presence of the FPP synthetase.

#### [Working Examples]

Examples of the preparation of the DNA sequence, plasmid, and transformant according to this invention will be described below, but the scope of this invention should not be restricted thereto.

#### Example 1

The experiment was carried out mainly following the procedures described in the above mentioned "Molecular cloning", "DNA Cloning" and "catalogue of Takara Shuzo Co., Ltd.". Enzymes used were mainly purchased from Takara Shuzo Co., Ltd.. Bacillus stearothermophilus used in a known bacterium deposited at American Type Culture Collection (ATCC). In this study, ATCC 10149 strain was used.

a) Preparation of Chromosomal DNA of *Bacillus stearothermophilus*

The strain was cultured in an LB medium (1% Trypton, 0.5% Yeast extract, 1% NaCl) and the cells were collected. After being suspended the cells in a lysis buffer, as described below, an amount of lysozyme (produced by Sigma, originating from avian albumen) was added so as to attain 10 mg/ml suspension. After lysis, 1/10 the amount of 1M Tris; HCl\* (pH 8.0), 1/10 the amount of 10% SDS, 1/50 the amount of 5 M NaCl were added. Proteinase K (produced by Sigma) was added so as to attain 10 mg/ml mixture, and the mixture was heated to 50 °C.

An equivalent amount of phenol was added, and the mixture was gently stirred, after which centrifugation was carried out to remove proteins. The centrifuged supernatant was transferred to a beaker by means of a wide mouthed pipet. After 2.5 times the amount of ethanol were overlaid, a chromosomal DNA was wounded by means of a glass rod. After being dissolved in a TE solution (10 mM Tris\*HCl, pH 8.0, 1 mM EDTA), the solution was treated with RNase A (produced by Sigma), with Proteinase K, and with phenol, after which ethanol was gently overlaid, and the chromosomal DNA was wound by means of a glass rod. After being washed with 70% ethanol, the chromosomal DNA was dissolved in the TE solution to be ready for the following experiment.

b) Preparation of Gene Library of *Bacillus stearothermophilus*

The chromosomal DNA was subjected to a partially digestion with restriction enzyme Sau3AI, and then to electrophoresis. An agarose containing DNA fragments from 2 kb.p. to 5 kb.p. was fractionated, and the DNA was extracted therefrom. The DNA was inserted into a BamHI restriction site on plasmid pUC118 (purchased from Takara Shuzo Co. Ltd.) using T4 DNA ligase to transform *Escherichia coli* JM109 strain. The library prepared as described above was used for screening.

c) Screening of Gene Encoding Isoprenoid Synthetase from the Library

The transformant from the library was cultured overnight in 30 ml of an LB medium containing 50 ug/ml of ampicillin, and the cells were collected. They were suspended in 3 ml of a sonic buffer, as described below, and the cells were lysed by sonication. The lysate were heated to 50 °C for 1 hour to inactivate the prenyltransferase originating from *Escherichia coli*, modified proteins originating *Escherichia coli* were centrifugally removed, and 600 µl of sample were used for the assay. The solution for the assay was reacted for 1 hour or 2 hour at 55 °C. The reacted solution was extracted with 1-butanol, and the radioactivity was determined by a liquid scintillation counter.

In the screening as described above, strong prenyltransferase was observed in pFE15.

Subsequently, the 1-butanol extract of JM109 having pFE15 which had been reacted as described above was analyzed by thin layer chromatography (TLC). As a result, the thus formed isoprenoid was identified as farnesyl pyrophosphate, and pFE15 was confirmed to contain a gene encoding a farnesyl pyrophosphate synthetase.

While the original *Escherichia coli* did not have any prenyltransferase having an activity at 55 °C, the *Escherichia coli* transformed with pFE15 had a prenyltransferase activity at 55 °C. The pFE15-coded prenyltransferase originating from *Bacillus stearothermophilus* was shown here to be very stable. The results are shown in Table 1. Here, the recombinant was shown to be effective for producing a stable farnesyl pyrophosphate.

The results of Determination of Prenyltransferase Activity (Radio-Activity of 1-Butanol Extract represented as dpm unit)		
JM109	pUC118/JM109	pFE15/JM109
$6.3 \times 10^2$	$7.8 \times 10^2$	$1.2 \times 10^5$

d) Production of pFE15-Deletion Mutant and Specification of FPP-Synthesizing Enzyme

pFE15 has an inserted gene of about 5 kb.p. and the restriction enzyme map is shown in Fig. 2. Of pFE15-deletion mutants, all the mutants containing the region from PstI to NruI were confirmed to possess an prenyltransferase activity. The plasmid pΔ1 in which about 2 kb.p. DNA fragment of PstI-NruI of pFE15

had been inserted in the SmaI site of pUC18 was observed to possess FPP synthetase activity. Whereas the activity remained when the portion of NruI of pΔ1 was deleted about 600 b.p., no activity remained when the PstI site was deleted about 100 b.p. Consequently, the gene encoding FPP synthetase was deduced to be at the PstI site slightly apart from NruI.

#### e) DNA sequencing of pΔ1

The DNA sequence of the deletion mutant produced with Exonuclease III or restriction enzymes was analyzed by an Applied Biosystem Model 373A florescent DNA sequencer. As a result, it was clarified to possess the base sequence depicted on Sequence No. 1 of the Sequence Listing and further it was deduced to possess an amino acid sequence appearing at the same time. When the deduced amino acid sequence was compared with the farnesyl pyrophosphate synthetase originating from *Escherichia coli* (J. Biochem. 108, page 995-100 (1990)), there was about 40% homology, confirmed to be a farnesyl pyrophosphate synthetase.

#### f) Production of Prenyltransferase Using lac Promoter

The plasmid pEX1 in which KpnI-pstI fragment of pΔ1 had been inserted in KpnI-PstI site of pUC19 was prepared (Fig. 1). Whereas in the pΔ1, the lac promoter and FPP synthetase gene have reversed direction to each other, in the pEX1, the lac promoter and FPP synthetase gene have the same direction. Subsequently, *Escherichia coli* JM109 was transformed with pEX1 and pΔ1, and the prenyltransferase activities were determined by the method described in section c). As a result, it was found that pEX1/JM109 had a prenyltransferase activity 15 times that of pΔ1/JM109, resulting in the productivity of FPP synthetase having been successfully improved with lac-promoter.

(Buffer: Lysis buffer)	
Sucrose	3 M
Tris* HCl (pH 8.0)	25 mM
EDTA	10 mM

(Buffer: Sonication buffer)	
Tris* HCl (pH 8.5)	50 mM
EDTA	1 mM
β-Mercaptoethanol	10 mM
PMSF	1 μg/ml
Phospholamidon	1 μg/ml

(Composition of reaction solution for assay (total 1 ml))	
[1- <sup>14</sup> C] isopentenyl pyrophosphate (produced by Amersham, corresponding to about $5.5 \times 10^5$ dpm)	25 nmol
Geranyl pyrophosphate	5 μmol
MgCl <sub>2</sub>	50 μmol
KF	5 μmol
β-Mercaptoethanol	50 μmol
Tris* HCl (pH 8.5)	50 μmol
Tris* HCl (pH 8.5)	50 μmol
Cell-free extract	600 μl

## SEQUENCE LISTING

SEQUENCE NUMBER: 1

5 SEQUENCE LENGTH: 894

SEQUENCE TYPE: Nucleic Acid

STRANDENESS: Double

TOPOLOGY: Linear

10 MOLECULE TYPE: Genomic DNA

SOURCE: Bacillus Stearothermophilus

SEQUENCE DESCRIPTION: SEQ ID NO: 1:

15 GTG GCG CAG CTT TCA GTT GAA CAG TTT CTC AAC GAG CAA AAA CAG 45  
Met Ala Met Leu Ser Val Glu Gln Phe Leu Asn Glu Gln Lys Gln  
5 10 15

20 GCG GTG GAA ACA GCG CTC TCC CGT TAT ATA GAG CGC TTA GAA GGG 90  
Ala Met Glu Thr Ala Leu Ser Arg Tyr Ile Glu Arg Leu Glu Gly  
20 25 30

25 CCG GCG AAG CTG AAA AAG GCG ATG GCG TAC TCA TTG GAG GCC GGC 135  
Pro Ala Lys Leu Lys Lys Ala Met Ala Tyr Ser Leu Glu Ala Gly  
35 40 45

30 GGC AAA CGA ATC CGT CCG TTG CTG CTT CTG TCC ACC GTT CGG GCG 180  
Gly Lys Arg Ile Arg Pro Leu Leu Leu Leu Ser Thr Val Arg Ala 60  
50 55 60

35 CTC GGC AAA GAC CCG GCG GTC GGA TTG CCC GTC GCC TGC GCG ATT 225  
Leu Gly Lys Asp Pro Ala Val Gly Leu Pro Val Ala Cys Ala Ile  
65 70 75

40 GAA ATG ATC CAT ACG TAC TCT TTG ATC CAT GAT GAT TTG CCG AGC 270  
Glu Met Ile His Thr Tyr Ser Leu Ile His Asp Asp Leu Pro Ser  
80 85 90

45 ATG GAC AAC GAT GAT TTG CGG CGC GGC AAG CCG ACG AAC CAT AAA 315  
Met Asp Asn Asp Asp Leu Arg Arg Gly Lys Pro Thr Asn His Lys  
95 100 105

50 GTG TTC GGC GAG GCG ATG GCC ATC TTG GCG GGG GAC GGG TTG TTG 360  
Val Phe Gly Glu Ala Met Ala Ile Leu Ala Gly Asp Gly Leu Leu  
110 115 120

ACG TAC GCG TTT CAA TTG ATC ACC GAA ATC GAC GAT GAG CGC ATC 405  
Thr Tyr Ala Phe Gln Leu Ile Thr Glu Ile Asp Asp Glu Arg Ile  
125 130 135

	CCT	CCT	TCC	GTC	CGG	CTT	CGG	CTC	ATC	GAA	CGG	CTG	GCG	AAA	GCG	450
	Pro	Pro	Ser	Val	Arg	Leu	Arg	Leu	Ile	Glu	Arg	Leu	Ala	Lys	Ala	
					140					145					150	
5	GCC	GGT	CCG	GAA	GGG	ATG	GTC	GCC	GGT	CAG	GCA	GCC	GAT	ATG	GAA	495
	Ala	Gly	Pro	Glu	Gly	Met	Val	Ala	Gly	Gln	Ala	Ala	Asp	Met	Glu	
					155					160					165	
10	GGA	GAG	GGG	AAA	ACG	CTG	ACG	CTT	TCG	GAG	CTC	GAA	TAC	ATT	CAT	540
	Gly	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	Glu	Tyr	Ile	His	
					170					175					180	
15	CGG	CAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	CAC	GCC	GGC	585
	Arg	His	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	His	Ala	Gly	
					185					190					195	
20	GCC	TTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	CTT	GAC	630
	Ala	Leu	Ile	Gly	Gly	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	Leu	Asp	
					200					205					210	
	GAA	TTC	GCC	GCC	CAT	CTA	GGC	CTT	GCC	TTT	CAA	ATT	CGC	GAT	GAT	675
	Glu	Phe	Ala	Ala	His	Leu	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	Asp	
25					215					220					225	
	ATT	CTC	GAT	ATT	GAA	GGG	GCA	GAA	GAA	AAA	ATC	GGC	AAG	CCG	GTC	720
	Ile	Leu	Asp	Ile	Glu	Gly	Ala	Glu	Glu	Lys	Ile	Gly	Lys	Pro	Val	
					230					235					240	
30	GGC	AGC	GAC	CAA	AGC	AAC	AAC	AAA	GCG	ACG	TAT	CCA	GCG	TTG	CTG	765
	Gly	Ser	Asp	Gln	Ser	Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	
					245					250					255	
35	TCG	CTT	GCC	GGC	GCG	AAG	GAA	AAG	TTG	GCG	TTC	CAT	ATC	GAG	GCG	810
	Ser	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Ala	Phe	His	Ile	Glu	Ala	
					260					265					270	
40	GCG	CAG	GCG	CAT	TTA	CGG	AAC	GCC	GAC	GTT	GAC	GGC	GCC	GCG	CTC	855
	Ala	Gln	Arg	His	Leu	Arg	Asn	Ala	Asp	Val	Asp	Gly	Ala	Ala	Leu	
					275					280					285	
	GCC	TAT	ATT	TGC	GAA	CTG	GTC	GCC	GCC	GCG	GAC	CAT	TAA			894
45	Ala	Tyr	Ile	Cys	Glu	Leu	Val	Ala	Ala	Arg	Asp	His	***			
					290					295						

A DNA sequence that encodes a stable farnesyl pyrophosphate synthetase and the use thereof are provided. DNA sequence that encodes a farnesyl pyrophosphate synthetase originating from Bacillus stearothermophilus is provided. By selecting Bacillus stearothermophilus as the gene origin of the synthetase, a production system for the synthetase, a production system for the synthetase particularly having thermal stability, can be constructed.

#### Claims

1. A DNA sequence that encodes an farnesyl pyrophosphate synthetase enzyme originating from Bacillus stearothermophilus.

2. The DNA sequence according to Claim 1, wherein said enzyme is an amino acid sequence depicted on Sequence No. 1 (SEQ ID NO: 1) of the Sequence Listing.
3. The DNA sequence according to claim 1, which comprises a DNA sequence depicted on Sequence No. 1 (SEQ ID NO: 1) of the Sequence Listing.
4. A recombinant vector comprising a DNA sequence of Claim 1 and a DNA sequence that has a function of regulating the expression of said DNA.
5. A microorganism cell having a gene encoding the farnesyl pyrophosphate synthetase transferred therein by the recombinant vector of Claim 4.
6. The microorganism according to Claim 5, which host cell is a microorganism cell belonging to the genus Escherichia.
7. A process for producing a farnesyl pyrophosphate synthetase active substance characterized by culturing the microorganism according to claim 5 or 6 in a nutrient medium to produce and accumulate the synthetase active substance in the cultures, and isolate the farnesyl pyrophosphate synthetase therefrom.
8. A process for preparation of a farnesyl pyrophosphate characterized by condensating a 3,3-dimethylallyl pyrophosphate or a geranyl pyrophosphate with an isopentenyl pyrophosphate in the presence of the farnesyl pyrophosphate synthetase active substance produced according to claim 7 to form the farnesyl pyrophosphate.

Fig.1

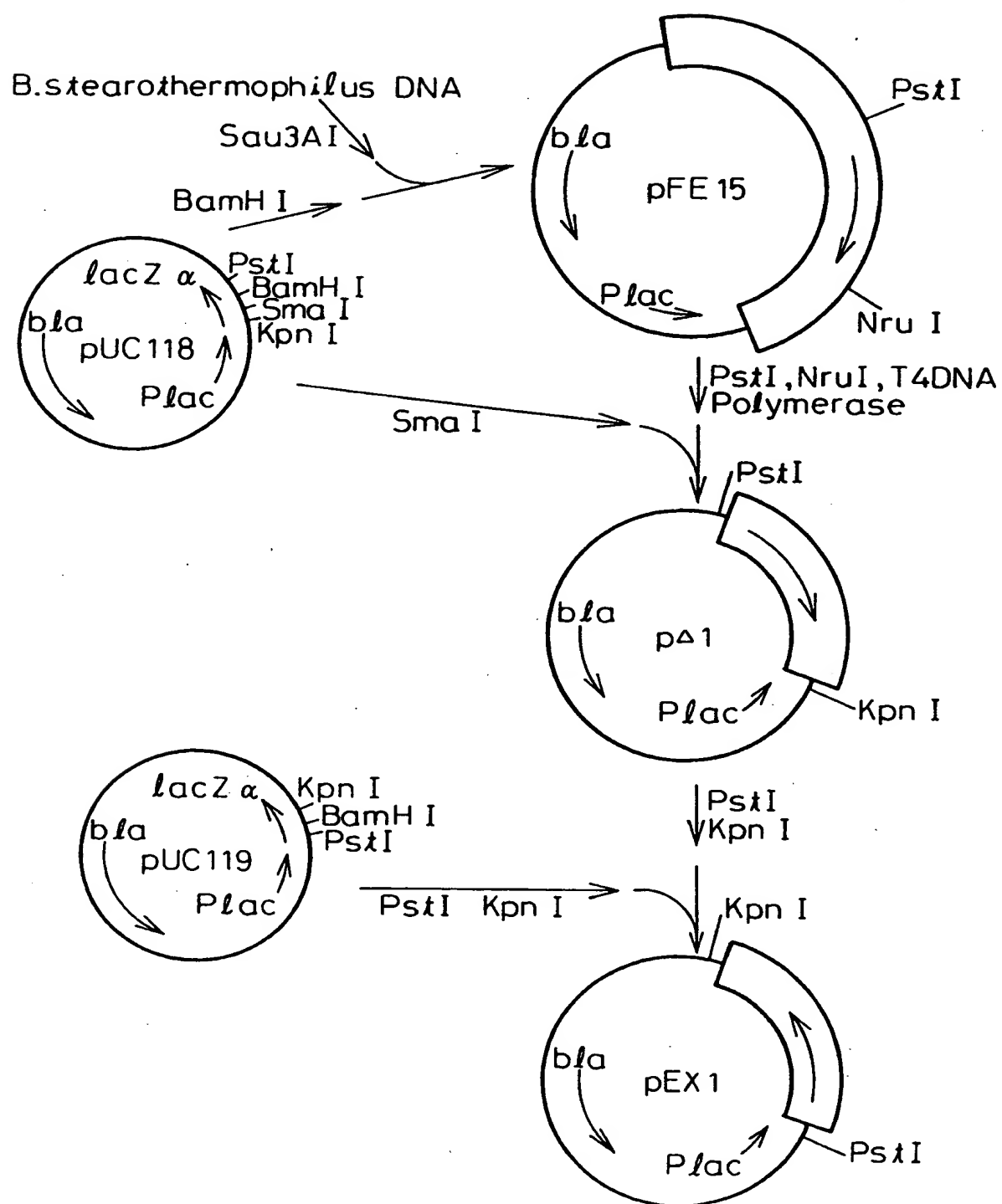


Fig.2

